

Conformational Analysis of (*R,S*)-4-Amido-2,4-dimethyl-butyr- ic Acid Derivatives

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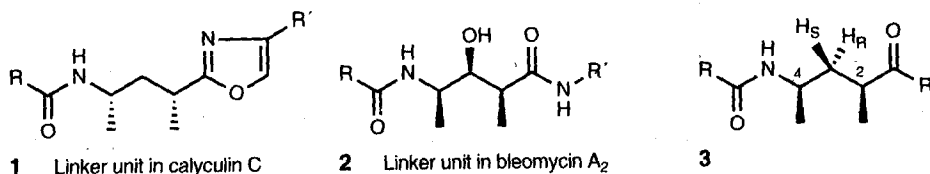
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Received 25 May 1999; accepted 21 June 1999

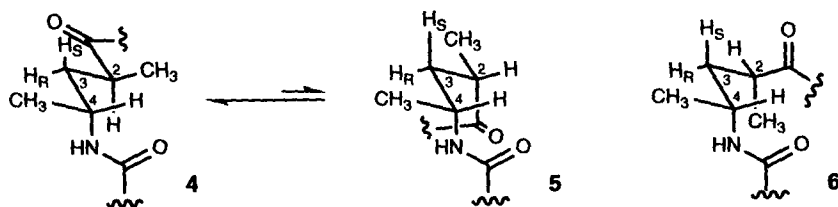
Keywords: Conformation; Hydrocarbon backbone

Abstract: NMR spectra of various *N*-acyl derivatives of (*R,S*)-4-amino-2,4-dimethyl-butyr-
ic acid show these compounds to be biconformational, populating the conformers
4 and 5 of the molecular backbone. Both predominant conformers have the amide
group gauche to the main chain. © 1999 Elsevier Science Ltd. All rights reserved.

The molecular backbone of biologically active compounds serves to position functional groups three-dimensionally in space for optimal interaction with the corresponding receptor. It is notable that nature uses flexible molecular backbones, which may have in many instances a preferred conformation. For example, in polypeptides nature combines α -amino acids as modules into flexible functional molecules with a pre-defined conformation. In search for non-natural analogs, oligomers of β -amino acids¹ have been studied with respect to distinct folding preferences. Recently, oligomers of γ -amino acids have been investigated in the same vein,² revealing distinct preferences for the formation of helical structures. In a few instances nature also relies on γ -amino acids as flexible linker units to confer a particular conformation to biologically active molecules. This is evidenced by the $^3J_{\text{H,H}}$ -NMR coupling constants along the backbone of calyculin C,³ cf. 1, or bleomycin A₂,⁴ cf. 2.



The linker unit in both natural products is a derivative of (*R,S*)-4-amino-2,4-dimethyl-butyr-
ic acid (3). MM3* calculations on 3 ($R = \text{CH}_3$, $R' = \text{OMe}$) suggest that two backbone-conformations 4, and 5
contribute predominantly to the local conformer population, whereas conformer 6 should be of sufficiently
higher energy (+ 7 kJ mol⁻¹ relative to 4) to be neglected.



Conformation 4 corresponds to that deduced for calyculin C³ or bleomycin A₂⁵ in solution from the NMR data. It corresponds as well to that found in the crystalline state for bleomycin A₂.⁶ We became interested to learn, whether this conformational preference is intrinsic to the backbone segment 3, or whether it is imposed to a large extent from the remainder of the molecule, i.e. the flanking groups R and R'. Knowledge of this sort is a prerequisite for the use of compounds 3 as spacer units in a modular approach to larger flexible molecules with defined shape.⁸

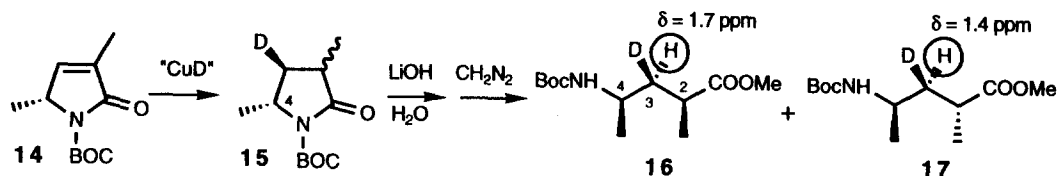
We therefore studied the conformational behaviour of a series of derivatives of 3 and related compounds⁹ by ¹H NMR spectroscopy (cf. table). The relevant vicinal H,H-coupling constants between the hydrogen atoms at C-2, C-3, and C-4 could not be determined in all cases completely, because of signal overlap or line broadening due to the presence of amide rotamers.

Table: NMR Coupling constants of (*R,S*)-4-Amido-2,4-dimethyl-Butyric Acid Derivatives

		Solvent	Temp (K)	δ (ppm)	C-3-H (pro- <i>R</i>)		C-3-H (pro- <i>S</i>)		
					³ J to H-2	³ J to H-4	δ (ppm)	³ J to H-2	³ J to H-4
1 a)		C ₆ D ₆	298	2.18	1.5	13.0	1.84	12.5	1.5
7 b)		CDCl ₃	298	1.94	6.2	9.6	1.68	6.7	n.d.
8		CDCl ₃	298	1.7	7	10	1.4	7 ^{c)}	n.d.
9		CDCl ₃	333	1.85	7.2	9.6	1.48	6.7 ^{c)}	4.8
10		CDCl ₃	333	1.87	7.2	9.8	1.48	n.d.	n.d.
11		CDCl ₃	298	1.89	8.0	9.9	1.49	n.d.	n.d.
12		CDCl ₃	328	1.86	7.6	10.3	1.49	6.5	3.9
13		C ₆ D ₆	298	1.11	6.6	8.7	1.01	7.7	n.d.

a) Values from ref. ³ ; b) values from ref. ⁷ ; c) Taken from the coupling pattern of the C-2-H signal; n.d. = not determined

In order to get more detailed information on the nature of the preferred conformer, the NMR-chemical shifts of the individual diastereotopic protons at C-3 of **8** were assigned by a stereospecific labelling procedure:



Starting from the pyrrolidinone **14**^{7,10} reduction with "copper deuteride"¹¹ provided a (1:1.5)-mixture of the C-2-epimeric pyrrolidinones **15**, the spectral data of which corresponded to those reported by Koskinen.^{7,10} We presume that the deuterium atom has been incorporated in this reduction *anti* to the C-4 methyl group of **15**. The pyrrolidinones were transformed into the esters **16** and **17** by treatment of **15** with LiOH followed by diazomethane. The esters **16** and **17** could then be separated by preparative HPLC. Comparison of the ^1H -NMR-spectra of **16** and of **8** showed that deuteration has led to the disappearance of the ^1H -NMR-signal at $\delta = 1.4$ ppm and had, moreover, simplified the coupling pattern of the C-3-H NMR signal at $\delta = 1.7$ ppm. This allowed an assignment of chemical shifts to the individual diastereotopic protons at C-3 in **8**. This assignment is in line with the one made for the diastereotopic protons in the corresponding segment **1** of calyculin C.³ In view of the consistency between **8** and **1**, the chemical shifts of the compounds **9** to **12** have been assigned by analogy.

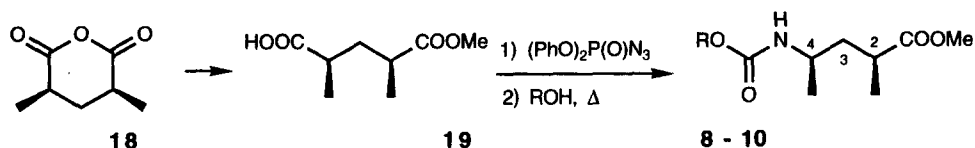
It is clear from the data in the table, that the conformational preference of the compounds **7** to **13** is lower than that of the corresponding segment **1** in calyculin C. It appears, however, that the same local conformer **4** is preferentially populated, albeit to a lesser extent. The consistently high $^3J_{\text{H,H}}$ -coupling constants between C-4-H and C-3- $\text{H}_{(\text{pro R})}$ indicate that the local conformation around the bond C-3/C-4 is fairly uniform. In turn, there is conformational heterogeneity regarding the C-2/C-3 bond. This is manifest from the low alteration of the values of the two C-2-H to C-3-H coupling constants. We therefore conclude that the 4-amino-2,4-dimethyl-butyrate segment in the compounds **7** to **13** is biconformational, with the conformations **4** and **5** populated to a similar extent. The backbone segment **3** present in compounds **7** to **13** has therefore a (moderate) intrinsic preference to populate a distinct conformation. This preference is increased in calyculin C or bleomycin A_2 by an (as not yet determined) effect of the groups flanking the segment **3**.

Acknowledgements: This work has been supported by the European Commission through the TMR network ERB FMRX CT 960011.

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The amides **11** and **12** were prepared by attaching the amido acid **10** to 2-chlorotrityl resin and coupling with the second amino acid by standard methods using *O*-benzotriazolyl-*N,N,N',N'*-tetramethyluronium tetrafluoroborate (TBTU) as coupling reagent.

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